

# Complete mitochondrial genome of *Lepidocephalichthys berdmorei* and its phylogenetic status within the family Cobitidae (Cypriniformes)

Min Zhou<sup>1</sup>, Cheng Wang<sup>2</sup>, Ziyue Xu<sup>1</sup>, Zhicun Peng<sup>1</sup>, Yang He<sup>1</sup>, Ying Wang<sup>1,3</sup>

1 Hubei Engineering Research Center for Protection and Utilization of Special Biological Resources in the Hanjiang River Basin, Jianghan University, Wuhan, China

2 State Key Laboratory of Freshwater Ecology and Biotechnology, Institute of Hydrobiology, Chinese Academy of Sciences, Wuhan, China

3 Academy of Plateau Science and Sustainability, Qinghai Normal University, Xining, China

Corresponding author: Ying Wang (xinyuanwangying@163.com)



## Abstract

In this study, the complete mitochondrial genome of *Lepidocephalichthys berdmorei* was first determined by the primer walking sequence method. The complete mitochondrial genome was 16,574 bp in length, including 13 protein-coding genes (PCGs), 22 transfer RNA (tRNA) genes, two ribosomal RNA (rRNA) genes, and a control region (D-loop). The gene arrangement pattern was identical to that of other teleosts. The overall base composition was 29.9% A, 28.5% T, 25.5% C, and 16.1% G, with an A+T bias of 58.4%. Furthermore, phylogenetic analyses were conducted based on 13 PCGs from the mitochondrial genomes of 18 cobitid species using three different methods (Neighbor-joining, Maximum likelihood, and Bayesian inference). All methods consistently showed that the four species of the genus *Lepidocephalichthys* form a monophyletic group. This study would provide effective molecular information for the *Lepidocephalichthys* species as well as novel genetic marker for the study of species identification.

Academic editor:  
Maria Elina Bichuette  
Received: 6 June 2024  
Accepted: 28 October 2024  
Published: 10 December 2024

ZooBank: <https://zoobank.org/DEE211BF-EF4D-42E2-AB1D-C82F8F8F4DC5>

Citation: Zhou M, Wang C, Xu Z, Peng Z, He Y, Wang Y (2024) Complete mitochondrial genome of *Lepidocephalichthys berdmorei* and its phylogenetic status within the family Cobitidae (Cypriniformes). ZooKeys 1221: 51–69. <https://doi.org/10.3897/zookeys.1221.129136>

Copyright: This is an open access article distributed under the terms of the [CC0 Public Domain Dedication](#).

## Introduction

*Lepidocephalichthys berdmorei* (Blyth 1860) belongs to the genus *Lepidocephalichthys* within the family Cobitidae, which is widely distributed in the Irrawaddy, Sittang, Salween, Chao Phraya, Mekong basins of Burma, Thailand, and China (Kottelat and Lim 1993). According to FishBase, there are approximately 25 valid species in the genus *Lepidocephalichthys* (Froese and Pauly 2024). The lack of reliable morphological characteristics, coupled with the widespread misapplication of names, has made it challenging to differentiate this species from its close relatives. For instance, the close resemblance in physical features between *L. thermalis* and *L. berdmorei* poses a significant challenge in morphological differentiation (Kottelat 2012). Therefore, molecular information is necessary for an additional method to delimit and identify species. *Lepidocephalichthys berdmorei* is a small-sized freshwater

fish species, that inhabits hill swift streams, and lakes with sandy and gravel bottoms (Kamei et al. 2023). In recent years, due to over-exploitation, damage to spawning beds, and construction of the hydroelectric dam in the Lancang River, the wild population size of *L. berdmorei* has declined dramatically (Buj et al. 2015; Zhang et al. 2019).

The mitochondrial genome (mtDNA) is a circular double-stranded molecule consisting of 13 PCGs, 22 tRNAs, two rRNAs, and a control region (D-loop) (Anderson et al. 1981; Boore 1999; Shen et al. 2020; Chu et al. 2022; Jia et al. 2023). Traditional morphological and biological approaches have focused on the ecological characteristics of populations and reproduction, with relatively little molecular research in the genus *Lepidocephalichthys* (Gohain and Deka 2017; Trif et al. 2022). Because of its limited recombination, highly conserved gene content, maternal inheritance and moderate evolutionary speed, mtDNA is now widely used to study population genetics, phylogeny, and species identification (Avise et al. 1987; Harrison 1989; Boore and Brown 1998; Ballard and Whitlock 2004; Galtier et al. 2009; Sureandiran et al. 2023). As proof, Wang et al. (2021) successfully identified fish species from the Xiangjiaba reservoir in Jinsha River using mitochondrial DNA barcoding. Goswami et al. (2022) characterized the genetic diversity of ten loaches from northeastern India based on sequence fragments of *cox1*, *cytb*, and *16S rRNA* genes; Zhang et al. (2023) demonstrated that the evolutionary position of *Rectoris luxiensis* (Wu et al. 1977) was consistent with traditional taxonomy through phylogenetic analysis of mitochondrial genomes. Currently, four mitochondrial genomes have been reported in NCBI databases, including *L. micropogon* (Blyth 1860), *L. guntea* (Hamilton 1822), *L. hasselti* (Valenciennes and Cuvier 1846), and *L. annandalei* (Chaudhuri 1912). Nevertheless, the complete mitochondrial genome of *L. berdmorei* has not been reported until now.

In this study, the complete mitochondrial genome of *L. berdmorei* was sequenced for the first time. The variation in tRNA length, position, and size of the control region, and the codon usage bias were analyzed. Subsequently, the 13 PCGs were concatenated and utilized, with those of other cobitids, to confirm the phylogenetic position of *L. berdmorei*. Therefore, these findings will provide valuable information and contribute to future species comparison and evolutionary research.

## Materials and methods

### Sample collection and DNA extraction

An adult individual of *L. berdmorei* was obtained in 2020 from the Mengla town, Xishuangbanna Dai Autonomous Prefecture, Yunnan Province, China (21°57'70"N, 101°60'54"E) (Suppl. material 1: fig. S1). Species were identified using the original morphological descriptions in the Fauna Sinica field guides (Chen 1998). After initial morphological identification, the specimen was deposited in the Animal Genetics Center of Jianghan University under the voucher number JHU202012029. A 40–50-mg fin clip was collected and preserved in 95% ethanol at 4 °C. Total genomic DNA was extracted from caudal fin tissue using the traditional phenol-chloroform method (Sambrook and Russell 2001).

## Mitogenome sequencing, assembly, and annotation

Eight pairs of primers (Suppl. material 1: table S1) were designed based on the mtDNA sequences of closely allied species. The PCR conditions were as follows: initial denaturation at 94 °C for 2 min, then 35 cycles of denaturation at 94 °C for 30 s, annealing at 55 °C for 30 s, and extension at 72 °C for 1 min, followed by the final extension at 72 °C for 10 min. All obtained fragments were quality-proofed and searched via BLAST in the NCBI database to confirm that the amplicon is the actual target sequence.

Sequences were assembled manually by the Seqman program using DNAstar v. 7.1 software (Burland 2000). The mitochondrial genome was annotated roughly following the procedure described before (Wang et al. 2011, 2018). The PCGs, rRNA genes, tRNA genes, and one control region of the mitochondrial genome were annotated by MitoAnnotator (<http://mitofish.aori.u-tokyo.ac.jp/annotation/input.html>) (Iwasaki et al. 2013). Their secondary structures of tRNAs were predicted by tRNAscan-SE (<http://lowelab.ucsc.edu/tRNAscan-SE/>; Lowe and Eddy 1997) and Forna (force-directed RNA) (Kerpedjiev et al. 2015).

The base composition and relative synonymous codon usage (RSCU) of the mitogenome were calculated and produced using PhyloSuite v. 1.2.3 (Zhang et al. 2020) and MAGA X (Kumar et al. 2018). The formulas to calculate the nucleotide composition of skew are as follows: AT-skew = (A – T) / (A + T) and GC-skew = (G – C) / (G + C) (Perna and Kocher 1995).

## Phylogenetic analyses

To verify the phylogenetic position of *L. berdmorei*, 17 mitogenome sequences from GenBank were retrieved (Suppl. material 1: table S2; Saitoh et al. 2006, 2010). The 13 PCGs for each species were concatenated and then aligned by program MAFFT using default settings (Katoh et al. 2002), and phylogenetic analyses were performed using Neighbor-joining (NJ), Maximum likelihood (ML), and Bayesian inference (BI) methods. To root the phylogenetic tree, *Syn-crossus beauforti* (Smith 1931) and *S. hymenophysa* (Bleeker 1852) from Botiidae were chosen as outgroups.

A NJ phylogenetic tree was constructed using MEGA 7 (Kumar et al. 2016) with 1,000 bootstrap replicates. The ML method was assembled in RAxML 7.0.3 (Stamatakis 2006), with 1,000 bootstrap replicates. GTR + F + I + G4 was selected as best-fit model according to Bayesian Information Criterions (BIC) estimated by ModelFinder (Kalyaanamoorthy et al. 2017). The BI phylogeny was carried out using MrBayes v. 3.2.7a (Ronquist et al. 2012) under the best-fit models with 5,000,000 generations in two runs of eight chains each.

## Abbreviations

Mitogenome, mitochondrial genome; **mtDNA**, mitochondrial DNA; **PCGs**, protein-coding genes; **tRNA**, transfer RNA; **rRNA**, ribosomal RNA; **atp6** and **atp8**, ATPase 6 and ATPase 8; **cox1–3**, cytochrome oxydasec subunits I–III; **cytb**, cytochrome b; **LA-PCR**, long and accurate polymerase chain reaction; **nd1–6**, NADH dehydrogenase subunits 1–6; **nd4l**, NADH dehydrogenase subunits 4L; **A+T**, A+T rich region; **RSCU**, relative synonymous codon usage; **trnA**, *tRNA*<sup>Ala</sup>;

**trnC**, *tRNA<sup>Cys</sup>*; **trnD**, *tRNA<sup>Asp</sup>*; **trnE**, *tRNA<sup>Glu</sup>*; **trnF**, *tRNA<sup>Phe</sup>*; **rrnS**, 12S rRNA; **rrnL**, 16S rRNA; **trnG**, *tRNA<sup>Gly</sup>*; **trnH**, *tRNA<sup>His</sup>*; **trnI**, *tRNA<sup>Ile</sup>*; **trnK**, *tRNA<sup>Lys</sup>*; **trnL1**, *tRNA<sup>Leu(TAA)</sup>*; **trnL2**, *tRNA<sup>Leu(TAG)</sup>*; **trnM**, *tRNA<sup>Met</sup>*; **trnN**, *tRNA<sup>Asn</sup>*; **trnP**, *tRNA<sup>Pro</sup>*; **trnQ**, *tRNA<sup>Gln</sup>*; **trnR**, *tRNA<sup>Arg</sup>*; **trnS1**, *tRNA<sup>Ser(TGA)</sup>*; **trnS2**, *tRNA<sup>Ser(GCT)</sup>*; **trnT**, *tRNA<sup>Thr</sup>*; **trnV**, *tRNA<sup>Val</sup>*; **trnW**, *tRNA<sup>Trp</sup>*; **trnY**, *tRNA<sup>Tyr</sup>*; **DHU**, Dihydrouracil; **NJ**, Neighbor-joining; **ML**, Maximum likelihood; **BI**, Bayesian inference.

## Results and discussion

### Mitogenome organization and nucleotide composition

The length of the complete mitochondrial genome of *L. berdmorei* is 16,574 bp (GenBank accession number: [OP651767](#)). The complete mitochondrial genome of *L. berdmorei* shares high similarity in gene arrangement, base composition, and codon usage pattern with those of other teleosts, indicating that the mitochondrial genome is highly conserved in evolution (Boore 1999; Taanman 1999; Broughton et al. 2001; Zou et al. 2019; Shen et al. 2020; Wang et al. 2020; Yu et al. 2021). The mitogenome is a circular double-stranded molecule with a highly conserved structure, consisting of 13 PCGs, 22 tRNA genes, two rRNA genes, and a control region (D-loop) (Fig. 1, Table 1).

The overall base composition is 29.9% for A, 16.1% for G, 25.5% for C, and 28.5% for T, which is consistent with the lowest frequency for G among the four bases in fish mitochondrial genomes, and revealing the A+T-rich content (58.4%) (Mayfield and McKenna 1978; Meyer 1993). Based on the analysis of nucleotide composition, this complete sequence exhibits a clear bias towards A and T (AT-skew = 0.02, GC-skew = -0.23) (Suppl. material 1: table S3). Both *L. berdmorei* and 58 species of Cobitidae exhibit an AT bias in their mitogenomes, but the A+T-rich content size varied among species, and it may be related to factors such as natural mutations and selection pressures during replication and transcription (Zhong et al. 2002; Yu et al. 2021). Hence, during the processes of replication and transcription, the asymmetry in nucleotide composition was used to infer the direction of gene orientation and replication (Francino and Ochman 1997; Frank and Lobry 1999; Satoh et al. 2016; Moeckel et al. 2023).

### Overlaps and non-coding intergenic spacers

Cobitidae mitogenomes range from 16,574 bp (*L. berdmorei*) to 16,646 bp (*Cobitis striata* (Ikeda, 1936)) in length (Suppl. material 1: table S2). With a few exceptions, the gene arrangements of fish mitogenomes are usually conserved (Anderson et al. 1981; Chang et al. 1994; Satoh et al. 2016; Chu et al. 2022). A typical feature in the mitochondrial genome of teleosts is the overlap of nucleotides between adjacent genes, suggesting that the size of mitochondrial DNA is very compact and economical, with potential kinetic advantages during the process of replication (Boore 1999; Curole and Kocher 1999; Taanman 1999; Wang et al. 2011; Satoh et al. 2016; Zou et al. 2017; Zou et al. 2018; Zhang et al. 2023). Similarly, in the *L. berdmorei* mitochondrial genome, there are overlaps and intervals of different lengths in all genes except for *trnF/rrnS*, *rrnS / trnV*, *trnV/rrnL*, *rrnL/trnL2*, *trnM/nd2*, *trnC/trnY*, *cox2/trnK*, *trnG/nd3*, *trnR/nd4l*, *trnH/trnS2*, *trnL2/nd5*, *nd6/trnE*, and *cytb/trnT*. They have the longest spacer in

trnN/trnC (30 bp) and the largest genetic overlap in *atp8/atp6* (10 bp) (Table 1). The length of the mitochondrial genome is related to the various overlaps and intergenic spacers between adjacent genes (Huang and Liu 2010). Interestingly, the presence of a specific 3 bp insertion (GCA) in the overlapping *atp8–atp6* motif of both *L. berdmorei* and other loaches compared to the conserved motif of 7 bp (ATGATAA) in other Cypriniformes fishes, suggests that this insertion is characteristic of loaches (Kanu et al. 2016; Wu et al. 2016; Yu et al. 2016; Yu et al. 2021). They may influence the expression of neighboring genes, regulate the normal operation of mitochondrial function, and participate in the process of mitochondrial genome replication and transmission (Boore 1999; Taanman 1999; D’Souza and Minczuk 2018).

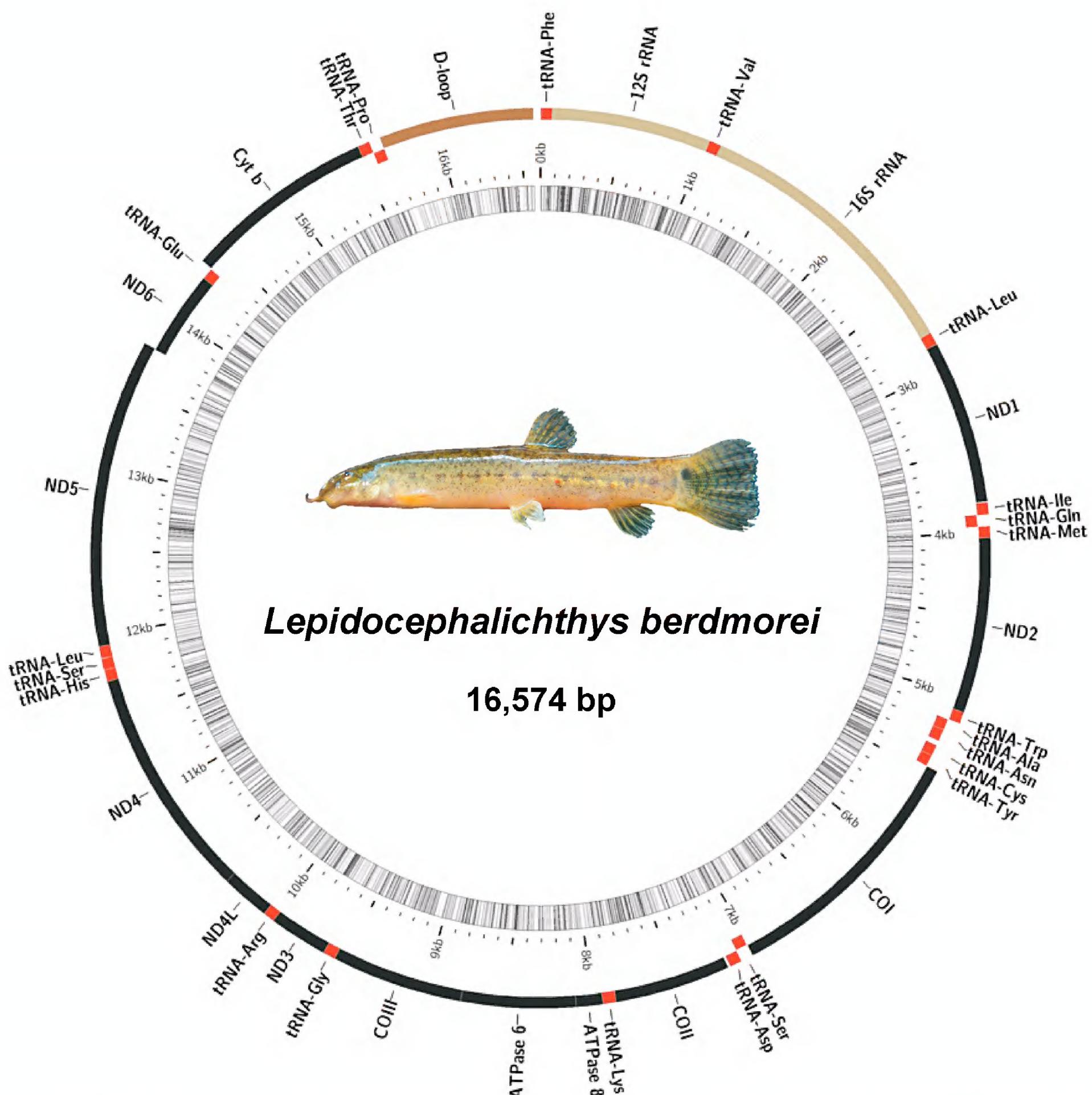


Figure 1. Gene map and organization of the mitochondrial genome of *Lepidocephalichthys berdmorei*. Photograph of *L. berdmorei* from <https://fishbase.se/summary/Lepidocephalichthys-berdmorei.html>.

**Table 1.** Organization of the mitochondrial genome of *Lepidocephalichthys berdmorei*.

Locus	Position		Size (bp)	Intergenic nucleotides <sup>a</sup>	Codon		Anti-codon	Strand <sup>b</sup>
	From	To			Start	Stop		
tRNA <sup>Phe(S)</sup>	1	69	69	0	–	–	GAA	H
12S rRNA	70	1019	950	0	–	–	–	H
tRNA <sup>Val(V)</sup>	1020	1091	72	0	–	–	TAC	H
16S rRNA	1092	2767	1676	0	–	–	–	H
tRNA <sup>Leu(TAA)(L1)</sup>	2768	2842	75	1	–	–	TAA	H
nd1	2844	3818	975	5	ATG	TAA	–	H
tRNA <sup>Ala(I)</sup>	3824	3895	72	-2	–	–	GAT	H
tRNA <sup>Gln(Q)</sup>	3894	3964	71	1	–	–	TTG	L
tRNA <sup>Met(M)</sup>	3966	4034	69	0	–	–	CAT	H
nd2	4035	5081	1047	-2	ATG	TAG	–	H
tRNA <sup>Trp(W)</sup>	5080	5148	69	2	–	–	TCA	H
tRNA <sup>Ala(A)</sup>	5151	5219	69	1	–	–	TGC	L
tRNA <sup>Asn(N)</sup>	5221	5293	73	30	–	–	GTT	L
tRNA <sup>Cys(C)</sup>	5324	5390	67	0	–	–	GCA	L
tRNA <sup>Tyr(Y)</sup>	5391	5459	69	1	–	–	GTA	L
cox1	5461	7011	1551	2	GTG	TAA	–	H
tRNA <sup>Ser(TGA)(S1)</sup>	7014	7084	71	1	–	–	TGA	L
tRNA <sup>Asp(D)</sup>	7086	7158	73	13	–	–	GTC	H
cox2	7172	7862	691	0	ATG	T	–	H
tRNA <sup>Lys(K)</sup>	7863	7938	76	1	–	–	TTT	H
atp8	7940	8107	168	-10	ATG	TAA	–	H
atp6	8098	8781	684	-1	ATG	TAA	–	H
cox3	8781	9566	786	-1	ATG	TAA	–	H
tRNA <sup>Gly(G)</sup>	9566	9638	73	0	–	–	TCC	H
nd3	9639	9989	351	-2	ATG	TAG	–	H
tRNA <sup>Arg(R)</sup>	9988	10056	69	0	–	–	TCG	H
nd4l	10057	10353	297	-7	ATG	TAA	–	H
nd4	10347	11729	1383	-1	ATG	TAG	–	H
tRNA <sup>His(H)</sup>	11729	11797	69	0	–	–	GTG	H
tRNA <sup>Ser(GCT)(S2)</sup>	11798	11866	69	1	–	–	GCT	H
tRNA <sup>Leu(TAG)(L2)</sup>	11868	11940	73	0	–	–	TAG	H
nd5	11941	13779	1839	-4	ATG	TAA	–	H
nd6	13776	14297	522	0	ATG	TAA	–	L
tRNA <sup>Glu(E)</sup>	14298	14366	69	5	–	–	TTC	L
cytb	14372	15512	1141	0	ATG	T	–	H
tRNA <sup>Thr(T)</sup>	15513	15584	72	-2	–	–	TGT	H
tRNA <sup>Pro(P)</sup>	15583	15652	70	0	–	–	TGG	L
D-loop	15653	16574	922	0	–	–	–	H

<sup>a</sup> Negative value indicates the overlapping sequences between adjacent genes.<sup>b</sup> H: heavy strand; L: light strand.

## PCGs and codon usage

The length of PCGs was 11,413 bp (68.86%) and it blanketed 7 NADH dehydrogenases (nd1–6 and nd4l), three cytochrome c oxidases (cox1–3), two ATPases (atp6 and atp8) and one cytochrome b (cytb). The size of PCGs ranged from nd4l (297 bp) to nd5 (1839 bp). As in other vertebrates, the nd6 and eight tRNA genes (tRNA<sup>Gln</sup>, tRNA<sup>Ala</sup>, tRNA<sup>Asn</sup>, tRNA<sup>Cys</sup>, tRNA<sup>Tyr</sup>, tRNA<sup>Ser</sup>, tRNA<sup>Pro</sup>, and tRNA<sup>Glu</sup>) are encoded on the light strand, and the others are encoded on the heavy strand (Fig. 1, Table 1) (Wen et al. 2017; Zou et al. 2017; Yu et al. 2021). In addition, the

bias of nucleotide composition was estimated (Suppl. material 1: table S3). All 13 PCGs showed a significant negative GC-skew. It may be that mutations in the replication process or adaptive evolution cause GC-skew. However, how to explain this unusual GC-skew needs further study.

Further analysis revealed that among 13 PCGs, most mitochondrial genes of *L. berdmorei* started with codon ATG, while only the *cox1* gene began with codon GTG. Unconventional start codons are a common phenomenon within the mitogenomes of fish (Zhang and Shen 2019; Yu et al. 2021). Eight of the PCGs are ended by TAA termination codons. The *nd2*, *nd3*, and *nd4* genes ended with TAG stop codons. The *cox2* and *cytb* use incomplete stop codons (T-) (Table 1). The relative synonymous codon usage (RSCU) denotes the differential usage of synonymous codons encoding the same amino acid. Essentially, the RSCU value was calculated by dividing the amino acids encoded by the same codons and their probability of appearing in the same codons (Sharp and Li 1986). The RSCUs of *L. berdmorei* mitogenome (Fig. 2, Table 2) show a clear preference for the usage of A and T. The total number of codons in the *L. berdmorei* mitochondrial genome is 5,524. After excluding the four stop codons (UAA(\*), UAG(\*), AGA(\*), AGG(\*)), among the 64 codons, 31 codons have an RSCU value greater than 1, indicating that these codons are prioritized more highly. For instance, six codons (UUU(L), UUG(L), CUU(L), CUC(L), CUA(L), CUG(L)) coded for leucine with preference for UUA. RSCU values for these six codons were 1.68, 0.64, 1.47, 0.65, 0.99 and 0.56, respectively. The most commonly used codon is UUU-Phe (F), followed by UUA-Leu2 (L), AAA-Lys (K), and AUU-Ile (I). The least used amino acids are Ala (GCG) and Arg (CGU). Our results show that the codon distribution is largely consistent with the mitogenomes of Cobitinae studied previously (Yu et al. 2021).

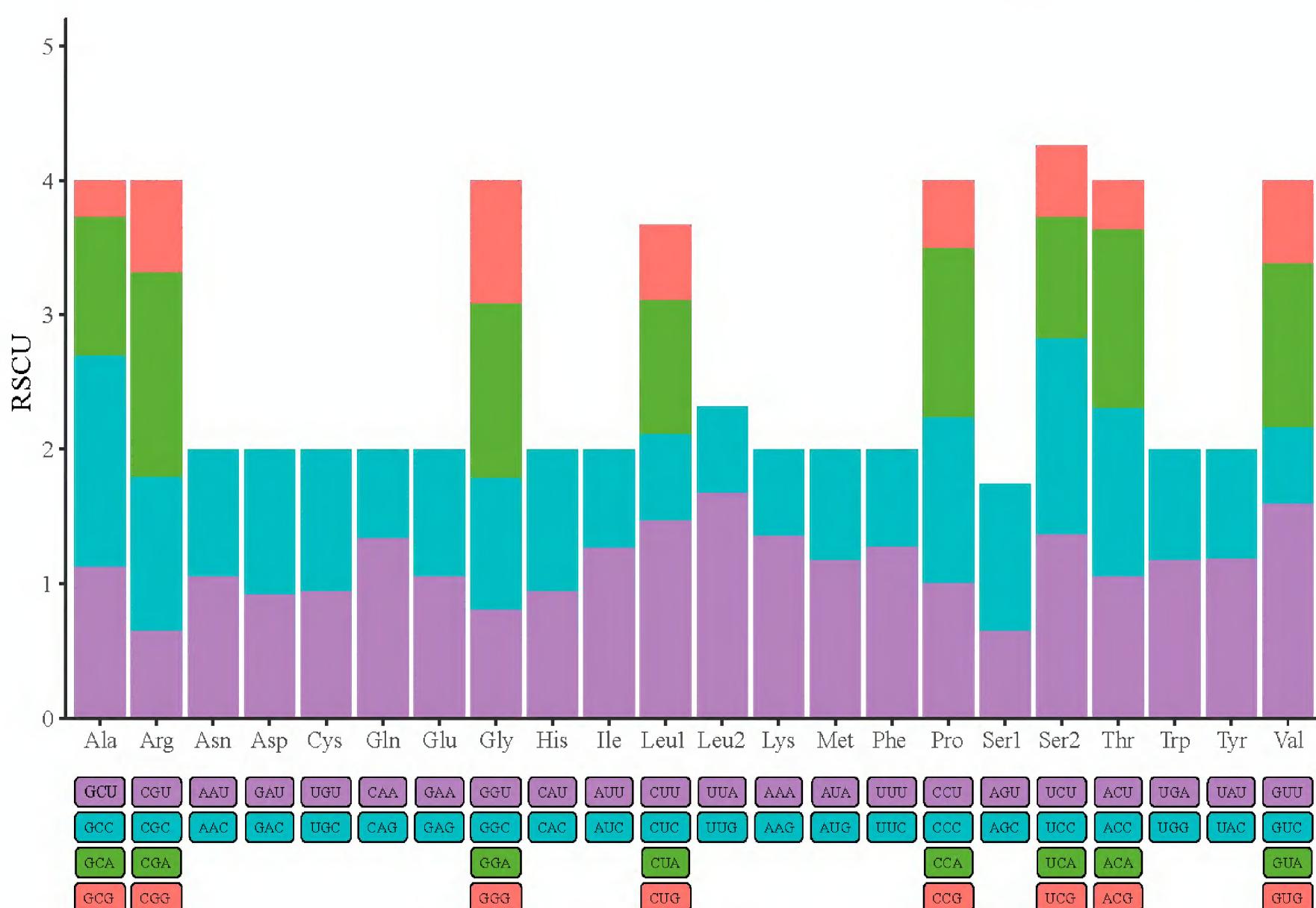


Figure 2. The relative synonymous codon usage (RSCU) of *L. berdmorei* mitogenome.

**Table 2.** Codon usage in the mitochondrial genome of *Lepidocephalichthys berdmorei*.

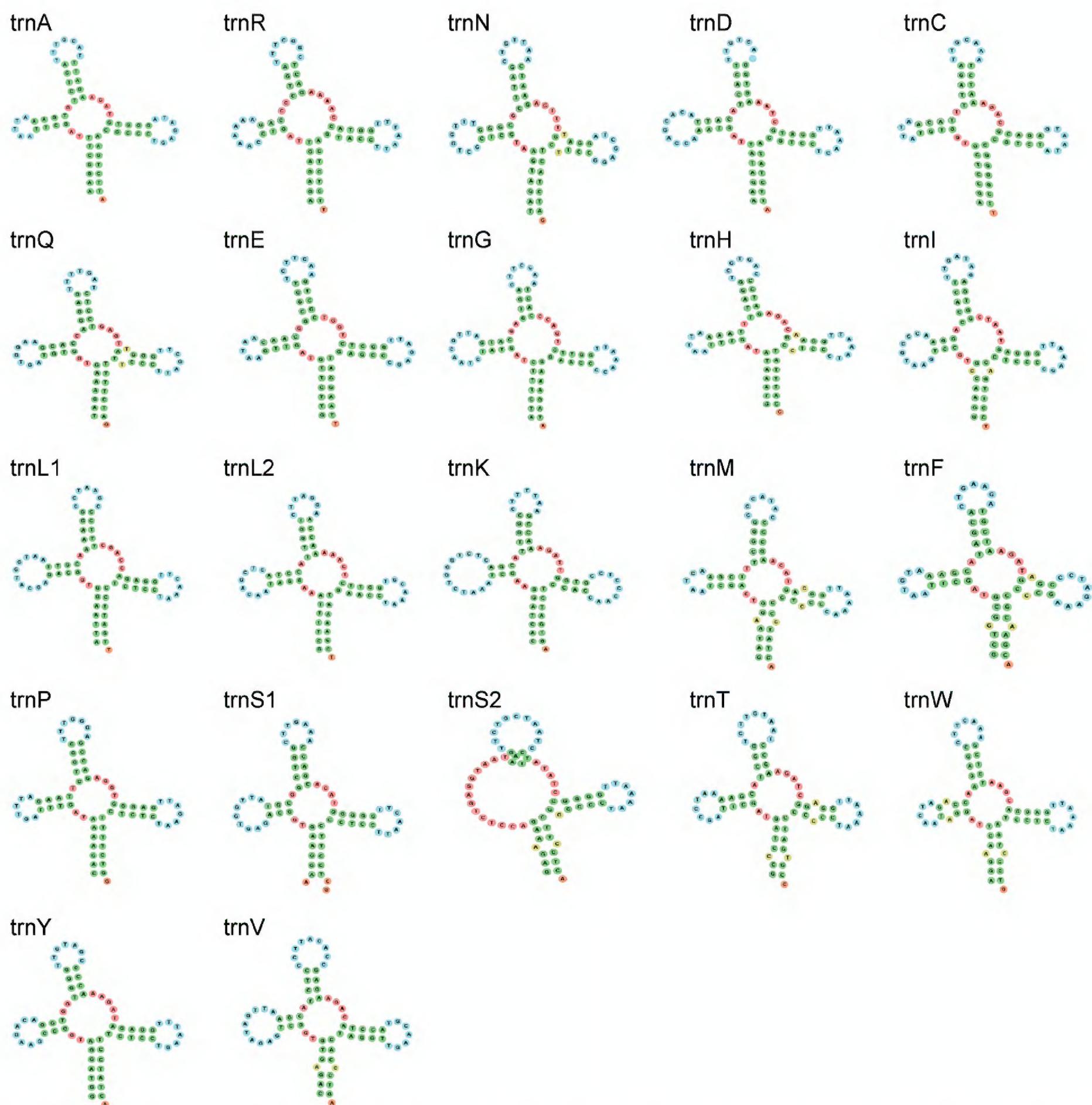
Codon	Count	RSCU									
UUU(F)	201	1.28	UCU(S)	103	1.37	UAU(Y)	118	1.19	UGU(C)	40	0.95
UUC(F)	113	0.72	UCC(S)	110	1.46	UAC(Y)	80	0.81	UGC(C)	44	1.05
UUA(L)	188	1.68	UCA(S)	68	0.9	UAA(*)	156	1.66	UGA(W)	75	1.18
UUG(L)	71	0.64	UCG(S)	40	0.53	UAG(*)	99	1.06	UGG(W)	52	0.82
CUU(L)	164	1.47	CCU(P)	110	1.01	CAU(H)	95	0.95	CGU(R)	21	0.65
CUC(L)	73	0.65	CCC(P)	134	1.23	CAC(H)	104	1.05	CGC(R)	37	1.15
CUA(L)	111	0.99	CCA(P)	137	1.26	CAA(Q)	156	1.34	CGA(R)	49	1.52
CUG(L)	63	0.56	CCG(P)	55	0.5	CAG(Q)	76	0.66	CGG(R)	22	0.68
AUU(I)	170	1.27	ACU(T)	102	1.06	AAU(N)	134	1.06	AGU(S)	49	0.65
AUC(I)	98	0.73	ACC(T)	120	1.25	AAC(N)	118	0.94	AGC(S)	82	1.09
AUA(M)	98	1.18	ACA(T)	128	1.33	AAA(K)	172	1.36	AGA(*)	75	0.8
AUG(M)	68	0.82	ACG(T)	35	0.36	AAG(K)	81	0.64	AGG(*)	45	0.48
GUU(V)	73	1.6	GCU(A)	82	1.13	GAU(D)	63	0.92	GGU(G)	44	0.81
GUC(V)	26	0.57	GCC(A)	114	1.57	GAC(D)	74	1.08	GGC(G)	53	0.98
GUA(V)	56	1.22	GCA(A)	75	1.03	GAA(E)	83	1.06	GGA(G)	70	1.3
GUG(V)	28	0.61	GCG(A)	20	0.27	GAG(E)	74	0.94	GGG(G)	49	0.91

### Transfer and ribosomal RNA genes

The complete mitogenome of *L. berdmorei* contains 22 tRNA genes with a size of 1,559 bp, 14 of which are located on the H-strand while the others are on the L-strand (Table 1). The 22 tRNA genes range from 67 bp to 76 bp in length, of which the shortest was *tRNA<sup>Cys</sup>* (67 bp) and the longest was *tRNA<sup>Lys</sup>* (76 bp). The color in Fig. 3 represents the type of tRNA structure in which the nucleotide is located. All tRNA genes have a typical cloverleaf secondary structure except *tRNA<sup>Ser(GCT)</sup>* lacking the Dihydrouracil (DHU) stem (Fig. 3). It is a common feature in many mitogenomes of metazoans, and can be integrated into ribosomes by adjusting its structure and function to fulfil its function of carrying and translocating amino acids (Watanabe et al. 2014; Liu et al. 2021; Xing et al. 2022).

The most prevalent non-Watson-Crick base pairs in the secondary structure of tRNAs are A-C (e.g., *trnI*, *trnH*, *trnM*, *trnV*, *trnS1*, *trnT*, *trnW*, and *trnF*), followed by T-T (*trnQ* and *trnN*), which are mostly located in the DHU, anticodon stems and acceptor (Fig. 3). And these mismatches may be modified by post-transcriptional editing processes without causing amino acid transport disorders (Tomita et al. 1996).

The lengths of 12S rRNA and 16S rRNA genes were 950 bp and 1,676 bp, which are located on the H strand (Table 1). They are bordered by *tRNA<sup>Phe</sup>* and *tRNA<sup>Leu(TAA)</sup>* and separated by *tRNA<sup>Val</sup>*. Both the lengths and base compositions of 12S rRNA and 16S rRNA are almost identical among the reported Cobitidae fishes (Kottelat and Lim 1993; Nalbant 1993; Yu et al. 2016; Shen et al. 2020; Chu et al. 2022; Ke et al. 2023). It shows a positive AT-skew (0.23) and a negative GC-skew (-0.07) (Suppl. material 1: table S3). Compared to entire mitochondrial genome, the 16S rRNA is a non-coding gene that evolves slowly, and it contains sufficient number of polymorphisms to distinguish species (Lakra et al. 2009; Sarri et al. 2014; Hossain et al. 2019). The 12S rRNA is also frequently considered as a DNA meta barcoding in fish identification and phylogenetic studies (Miya et al. 2015).



**Figure 3.** Putative secondary structure of tRNAs. Stems (typically helical) are shown in green, multiple loops (junctions) are shown in red, interior loops are shown in yellow, hairpin loops are shown in blue, and 5' and 3' unpaired regions are shown in orange.

### Control region

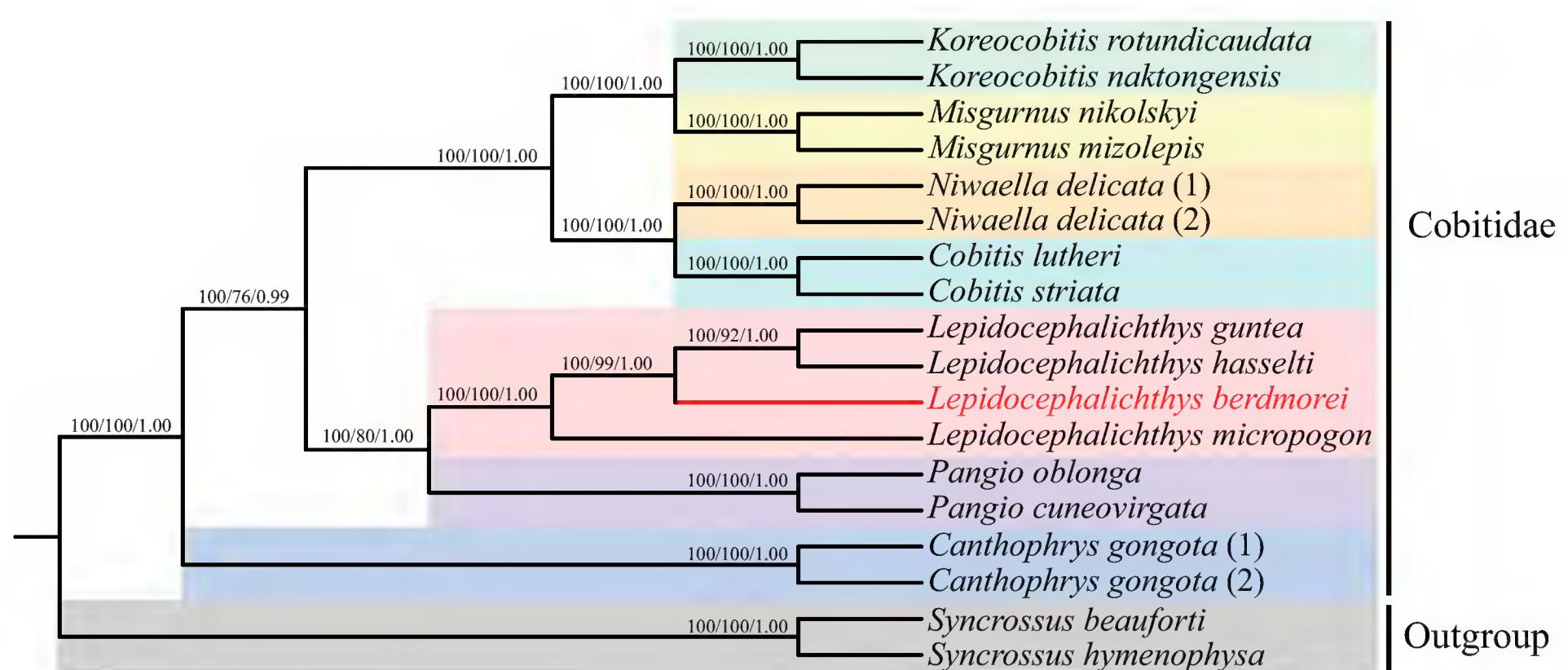
The only large control region of *L. berdmorei* mitogenome is the D-loop, located between the *tRNA<sup>Pro</sup>* and *tRNA<sup>Phe</sup>*, with a length of 922 bp (Fig. 1, Table 1). It plays a role in the regulation of replication and transcription and is the most rapidly evolving and changing region of the mitochondrial genome (Clayton 1982, 1991; Shadel and Clayton 1997; Zhou et al. 2014; Gao et al. 2023). The A+T-rich content of the *L. berdmorei* D-loop region is 66.27%, which is higher than the average value of the whole mitogenome (58.43%) and 13 PCGs (56.11–61.07%) (Suppl. material 1: table S3), as found in other vertebrates (Brown et al. 1986; Saccone et al. 1987; Zou et al. 2017; Ke et al. 2023).

In addition to gene duplication and insertion/deletion events, the main cause of mitochondrial genome size variation is differences in control region length (Mignotte et al. 1990; Lee et al. 1995; Pereira 2000; Minhas et al. 2023). Previous studies have demonstrated that tandem repeat sequences are prevalent in the D-loop of teleost lineage (Lee et al. 1995; Nicholls and Minczuk 2014; Jemt et al. 2015; Xu et al. 2016; Ke et al. 2023). It is worth noting that the copy number not only varies between species, but also among individuals within the same species (Norman et al. 1994; Lunt et al. 1998; Boore 1999; Xu et al. 2021). Thus, compared with the complex and large eukaryotic genome, the mitochondrial genome is simple in structure with shorter sequences, contains both conserved and highly variable regions, and can be used for taxonomic identification of species at different levels of evolution (Pereira et al. 2008; Jamandre et al. 2014; Nicholls and Minczuk 2014; Jemt et al. 2015; D'Souza and Minczuk 2018). Nevertheless, multiple duplicate regions have been found in some species that may adversely affect PCR amplification, sequencing, or both (Singh et al. 2008; Cadahía et al. 2009). As a result, researchers have avoided using this region for phylogenetic purposes, focusing instead on *rRNA* or *PCGs* (Slechtová et al. 2008; Wang et al. 2021; Sureandiran et al. 2023; Zhang et al. 2023).

## Phylogenetic analysis

Cobitidae belongs to Osteichthyes, Cyprinidformes, and has three subfamilies: Nemacheilinae, Botiinae and Cobitinae (Hora 1932; Nalbant 1993; Tang et al. 2005; Slechtová et al. 2008; Chu et al. 2022). Sawada (1982) proposed a phylogeny of the Cobitoidea (limited to loaches) as (Botiinae + Cobitinae) + (Nemacheilinae + Homalopterinae) based on 52 osteological characters. Nevertheless, due to their morphological similarity and frequent overlap, differentiating species within Cobitidae based solely on morphology is a challenging endeavor (Kottelat and Lim 1993; Nalbant 1993; Shen et al. 2020; Ke et al. 2023). In order to determine the phylogenetic status of *L. berdmorei* in the family Cobitidae, 17 complete mitochondrial genomes from the GenBank database were selected to reconstruct phylogenetic trees. Based on the 13 PSGs concatenated dataset, the NJ, ML and BI phylogenies generated identical topology with high bootstrap support and posterior probability values, respectively (Fig. 4). All trees presented two major clades corresponding to the outgroup. *Canthophrys* is located at the base of the phylogenetic tree. Our results are generally consistent with the traditional morphological classification and recent molecular studies (Hora 1932; Slechtová et al. 2008; Sudasinghe et al. 2024).

Firstly, the phylogenetic tree revealed that *L. guntea*, *L. hasselti*, and *L. berdmorei* clustered as a monophyletic clade, followed by a clade with *L. micropogon* with high bootstrap support. Secondly, the genus *Lepidocephalichthys* and *Pangio* which formed a sister branch with high bootstrap support and posterior probability values, which was consistent with the previous study (Slechtová et al. 2008; Yu et al. 2021). Notably, Slechtová et al. (2008) found that the genera *Lepidocephalichthys* and *Pangio* were considered as a sister group in the RAG-1 phylogeny; but this relationship was not supported by the *cytb* dataset. Meanwhile, based on *cyt b* and RAG-1 datasets, these four genera of Cobitidae (*Cobitis*, *Niwaella*, *Misgurnus*, and *Koreocobitis*) form a distinct monophyletic group (Slechtová et al. 2008). Generally, from the phylogenetic tree of genetic evolution, the evolutionary status of *L. berdmorei* was defined.



**Figure 4.** Phylogenetic tree of Cobitidae and two outgroups based on the NJ, ML and BI analysis of 13 concatenated protein-coding genes. Tree topologies produced by NJ, ML methods, and BI analysis were equivalent. The numbers at the nodes represent bootstrap support values for NJ and ML analyses and Bayesian posterior probability, sequentially, and the red branch represents the species in this study.

## Conclusions

In conclusion, the complete mitochondrial DNA sequence of *L. berdmorei* is determined for the first time by the primer walking sequence method. The mitogenome is 16,574 bp in length, and encodes all of the 37 genes that are typical for Cobitidae fish. We compared mtDNA from *L. berdmorei* with that of other teleost and analyzed mitogenome composition, PCGs, and codon usage, transfer and ribosomal RNA genes, and noncoding regions (control region, intergenic spacers). The generated phylogenetic trees yielded convincing evidence that the genus *Lepidocephalichthys* formed a monophyletic group. These findings will provide new insights into better understanding the phylogenetic status of this intriguing and ecologically important group.

## Acknowledgements

This work was supported by grants from the National Natural Science Foundation of China (31702016), and Jianghan University Research Project Funding Plan (2023KJZX42). We greatly appreciate the valuable suggestions of two anonymous referees.

## Additional information

### Conflict of interest

The authors have declared that no competing interests exist.

### Ethical statement

All animal protocols have been reviewed and approved by the experimental animal welfare and ethics review committee of Jianghan University, Qinghai Normal University, and Chinese Academy of Sciences.

## Funding

This work was supported by the National Natural Science Foundation of China (Grant number 31702016) and Jianghan University Research Project Funding Plan (Grant number 2023KJZX42).

## Author contributions

Ying Wang contributed to the concept and design of the study. Sample collection and preparation of materials were done by Cheng Wang and Zhicun Peng. Ziyue Xu, Yang He and Min Zhou performed the data analysis and interpretation, and Min Zhou wrote the first draft of the manuscript. Ying Wang revised this manuscript. All authors read and revised the manuscript and approved the final version. All authors agree to be accountable for all aspects of the manuscript.

## Author ORCIDs

Ying Wang  <https://orcid.org/0000-0002-8222-7510>

## Data availability

Genome sequence data that support the findings of this study are openly available from the GenBank at <https://www.ncbi.nlm.nih.gov/>, under accession No. [OP651767](#).

## References

Anderson S, Bankier AT, Barrell BG, de Bruijn MH, Coulson AR, Drouin J, Eperon IC, Nierlich DP, Roe BA, Sanger F, Schreier PH, Smith AJ, Staden R, Young IG (1981) Sequence and organization of the human mitochondrial genome. *Nature* 290(5806): 457–465. <https://doi.org/10.1038/290457a0>

Avise JC, Arnold J, Ball RM, Bermingham E, Lamb T, Neigel JE, Reeb CA, Saunders NC (1987) Intraspecific phylogeography: the mitochondrial DNA bridge between population genetics and systematics. *Annual Review of Ecology and Systematics* 18: 489–522. <https://doi.org/10.1146/annurev.es.18.110187.002421>

Ballard JWO, Whitlock MC (2004) The incomplete natural history of mitochondria. *Molecular Ecology* 13(4): 729–744. <https://doi.org/10.1046/j.1365-294X.2003.02063.x>

Bleeker P (1852) Diagnostische beschrijvingen van nieuwe of weinig bekende vischsoorten van Sumatra. Tiental I–IV. *Natuurkundig Tijdschrift Voor Nederlandsch Indië* 3: 569–608. <https://www.biodiversitylibrary.org/part/218337>

Blyth E (1860) Report on some fishes received chiefly from the Sitang River and its tributary streams. *The Journal of the Asiatic Society of Bengal* 29: 138–174. <https://biostor.org/reference/235393>

Boore JL (1999) Animal mitochondrial genomes. *Nucleic Acids Research* 27(8): 1767–1780. <https://doi.org/10.1093/nar/27.8.1767>

Boore JL, Brown WM (1998) Big trees from little genomes: mitochondrial gene order as a phylogenetic tool. *Current Opinion in Genetics & Development* 8(6): 668–674. [https://doi.org/10.1016/S0959-437X\(98\)80035-X](https://doi.org/10.1016/S0959-437X(98)80035-X)

Broughton RE, Milam JE, Roe BA (2001) The complete sequence of the zebrafish (*Danio rerio*) mitochondrial genome and evolutionary patterns in vertebrate mitochondrial DNA. *Genome Research* 11(11): 1958–1967. <https://doi.org/10.1101/gr.156801>

Brown GG, Gadaleta G, Pepe G, Saccone C, Sbisà E (1986) Structural conservation and variation in the D-loop-containing region of vertebrate mitochondrial DNA. *Journal of Molecular Biology* 192(3): 503–511. [https://doi.org/10.1016/0022-2836\(86\)90272-X](https://doi.org/10.1016/0022-2836(86)90272-X)

Buj I, Ćaleta M, Marčić Z, Šanda R, Vukić J, Mrakovčić M (2015) Different histories, different destinies-impact of evolutionary history and population genetic structure on extinction risk of the adriatic spined loaches (genus *Cobitis*; Cypriniformes, Actinopterygii). PLoS ONE 10(7): e0131580. <https://doi.org/10.1371/journal.pone.0131580>

Burland TG (2000) DNASTAR's Lasergene sequence analysis software. Methods in Molecular Biology 132: 71–91. <https://doi.org/10.1385/1-59259-192-2:71>

Cadahía L, Pinsker W, Negro JJ, Pavlicev M, Urios V, Haring E (2009) Repeated sequence homogenization between the control and pseudo-control regions in the mitochondrial genomes of the subfamily Aquilinae. Journal of Experimental Zoology Part B: Molecular and Developmental Evolution 312B(3): 171–185. <https://doi.org/10.1002/jez.b.21282>

Chang YS, Huang FL, Lo TB (1994) The complete nucleotide sequence and gene organization of carp (*Cyprinus carpio*) mitochondrial genome. Journal of Molecular Evolution 38(2): 138–155. <https://doi.org/10.1007/BF00166161>

Chaudhuri, BL (1912) Descriptions of some new species of freshwater fishes from north India. Records of the Zoological Survey of India 7(5): 437–443. <https://doi.org/10.26515/rzsi/v7/i5/1912/163113>

Chen YY (1998) *Fauna Sinica*, Osteichthyes, Cypriniformes (II). Science Press, Beijing, 531 pp.

Chu TJ, Yang N, Liu K (2022) Complete mitochondrial genome identification of *Niwaella nigrolinea* (Cypriniformes: Cobitidae). Mitochondrial DNA. Part B, Resources 7(11): 2018–2020. <https://doi.org/10.1080/23802359.2022.2149247>

Clayton DA (1982) Replication of animal mitochondrial DNA. Cell 28(4): 693–705. [https://doi.org/10.1016/0092-8674\(82\)90049-6](https://doi.org/10.1016/0092-8674(82)90049-6)

Clayton DA (1991) Nuclear gadgets in mitochondrial DNA replication and transcription. Trends in Biochemical Sciences 16(3): 107–111. [https://doi.org/10.1016/0968-0004\(91\)90043-U](https://doi.org/10.1016/0968-0004(91)90043-U)

Curole JP, Kocher TD (1999) Mitogenomics: digging deeper with complete mitochondrial genomes. Trends in Ecology & Evolution 14(10): 394–398. [https://doi.org/10.1016/S0169-5347\(99\)01660-2](https://doi.org/10.1016/S0169-5347(99)01660-2)

D'Souza AR, Minczuk M (2018) Mitochondrial transcription and translation: overview. Essays in Biochemistry 62(3): 309–320. <https://doi.org/10.1042/EBC20170102>

Francino MP, Ochman H (1997) Strand asymmetries in DNA evolution. Trends in Genetics 13(6): 240–245. [https://doi.org/10.1016/S0168-9525\(97\)01118-9](https://doi.org/10.1016/S0168-9525(97)01118-9)

Frank AC, Lobry JR (1999) Asymmetric substitution patterns: a review of possible underlying mutational or selective mechanisms. Gene 238(1): 65–77. [https://doi.org/10.1016/S0378-1119\(99\)00297-8](https://doi.org/10.1016/S0378-1119(99)00297-8)

Froese R, Pauly D (2024) FishBase. World Wide Web electronic publication. <http://www.fishbase.org> [accessed May 2024]

Galtier N, Nabholz B, Glémin S, Hurst GDD (2009) Mitochondrial DNA as a marker of molecular diversity: a reappraisal. Molecular Ecology 18(22): 4541–4550. <https://doi.org/10.1111/j.1365-294X.2009.04380.x>

Gao Y, Zhang J, Wang Q, Liu Q, Tang B (2023) The complete mitochondrial genome of box tree moth *Cydalima perspectalis* and insights into phylogenetics in Pyraloidea. Animals: An Open Access Journal from MDPI 13(6): 1045. <https://doi.org/10.3390/ani13061045>

Gohain AB, Deka P (2017) Length-weight relationship and relative condition factor of *Lepidocephalichthys Guntea* (Hamilton, 1822) of Ghati Beel of Dhemaji district of Assam, India. International Journal of Fisheries and Aquatic Studies 5(2): 514–517.

Goswami M, Pavan-Kumar A, Patil G, George T, Nath R, Bhuyan RN, Siva C, Laskar MA, Sumer S (2022) Molecular identification of ornamental loaches (Cypriniformes, Co-

bitoidei) of North East India using mitochondrial genes. *Animal Gene* 26: 200136. <https://doi.org/10.1016/j.angen.2022.200136>

Hamilton F (1822) An account of the fishes found in the river Ganges and its branches. Printed for A. Constable and company, Edinburgh, 405 pp. <https://doi.org/10.5962/bhl.title.6897>

Harrison RG (1989) Animal mitochondrial DNA as a genetic marker in population and evolutionary biology. *Trends in Ecology & Evolution* 4(1): 6–11. [https://doi.org/10.1016/0169-5347\(89\)90006-2](https://doi.org/10.1016/0169-5347(89)90006-2)

Hora SL (1932) Classification, bionomics and evolution of homalopteridae fishes. *Zoological Survey of India*, Calcutta, 263–330 pp.

Hossain MAM, Uddin SMK, Chowdhury ZZ, Sultana S, Johan MR, Rohman A, Erwanto Y, Ali ME (2019) Universal mitochondrial 16s rRNA biomarker for mini-barcode to identify fish species in Malaysian fish products. *Food Additives & Contaminants: Part A: Chemistry, Analysis, Control, Exposure & Risk Assessment* 36(4): 493–506. <https://doi.org/10.1080/19440049.2019.1580389>

Huang Z, Liu N (2010) Advances in mitochondrial genome size variation in animals. *Life Science Research* 14(02):166–171. <https://doi.org/10.3724/SP.J.1238.2010.00512>

Ikeda H (1936) On the sexual dimorphism and the taxonomical status of some Japanese loaches (I). *Zoological Magazine*, 48: 983–994. <https://doi.org/10.34435/zm002513>

Iwasaki W, Fukunaga T, Isagozawa R, Yamada K, Maeda Y, Satoh TP, Sado T, Mabuchi K, Takeshima H, Miya M, Nishida M (2013) MitoFish and MitoAnnotator: a mitochondrial genome database of fish with an accurate and automatic annotation pipeline. *Molecular Biology and Evolution* 30(11): 2531–2540. <https://doi.org/10.1093/molbev/mst141>

Jamandre BW, Durand JD, Tzeng WN (2014) High sequence variations in mitochondrial DNA control region among worldwide populations of flathead mullet *Mugil cephalus*. *International Journal of Zoology* 2014(2014): 1–9. <https://doi.org/10.1155/2014/564105>

Jemt E, Persson Ö, Shi Y, Mehmedovic M, Uhler JP, Dávila López M, Freyer C, Gustafsson CM, Samuelsson T, Falkenberg M (2015) Regulation of DNA replication at the end of the mitochondrial D-loop involves the helicase TWINKLE and a conserved sequence element. *Nucleic Acids Research* 43(19): 9262–9275. <https://doi.org/10.1093/nar/gkv804>

Jia W, Wei J, Niu M, Zhang H, Zhao Q (2023) The complete mitochondrial genome of *Aeschrocoris tuberculatus* and *A. ceylonicus* (Hemiptera, Pentatomidae) and its phylogenetic implications. *ZooKeys* 1160: 145–167. <https://doi.org/10.3897/zookeys.1160.100818>

Kalyaanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermiin LS (2017) ModelFinder: fast model selection for accurate phylogenetic estimates. *Nature Methods* 14(6): 587–589. <https://doi.org/10.1038/nmeth.4285>

Kamei M, Munilkumar S, Basudha C, Kamei M, Munilkumar S, Basudha C, Dasgupta S, Sawant PB, Romen Mangang W (2023) Breeding and larval rearing of juvenile of Burmese loach, *Lepidocephalichthys berdmorei* (Blyth, 1860): A new candidate species for aquaculture. *Journal of Experimental Zoology India* 26(1): 11–17. <https://connectjournals.com03895.2023.26.11>

Kanu UC, Zhao G, Xie P, Li Y, Lei D, Niu J, Ma X (2016) The complete mtDNA genome of *Triplophysa strauchi* (Cypriniformes, Balitoridae, Cobitoidea): genome characterization and phylogenetic analysis. *Mitochondrial DNA. Part A, DNA Mapping, Sequencing, and Analysis* 27(4): 2637–2638. <https://doi.org/10.3109/19401736.2015.1041131>

Katoh K, Misawa K, Kuma K, Miyata T (2002) MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. *Nucleic Acids Research* 30(14): 3059–3066. <https://doi.org/10.1093/nar/gkf436>

Ke Z, Zhou K, Hou M, Luo H, Li Z, Pan X, Zhou J, Jing T, Ye H (2023) Characterization of the complete mitochondrial genome of the elongate loach and its phylogenetic implications in Cobitidae. *Animals: An Open Access Journal from MDPI* 13(24): 3841. <https://doi.org/10.3390/ani13243841>

Kerpedjiev P, Hammer S, Hofacker IL (2015) Forna (force-directed RNA): Simple and effective online RNA secondary structure diagrams. *Bioinformatics* (Oxford, England) 31(20): 3377–3379. <https://doi.org/10.1093/bioinformatics/btv372>

Kottelat M (2012) Conspectus cobitidum: an inventory of the loaches of the world (Teleostei: Cypriniformes: Cobitoidei). *The Raffles Bulletin of Zoology Suppl* 26: 1–199.

Kottelat M, Lim KKP (1993) A synopsis of the Malayan species of *Lepidocephalichthys*, with descriptions of two new species (Teleostei: Cobitidae). *The Raffles Bulletin of Zoology* 40(2): 201–220.

Kumar S, Stecher G, Tamura K (2016) MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution* 33(7): 1870–1874. <https://doi.org/10.1093/molbev/msw054>

Kumar S, Stecher G, Li M, Knyaz C, Tamura K (2018) MEGA X: Molecular evolutionary genetics analysis across computing platforms. *Molecular Biology and Evolution* 35(6): 1547–1549. <https://doi.org/10.1093/molbev/msy096>

Lakra WS, Goswami M, Gopalakrishnan A (2009) Molecular identification and phylogenetic relationships of seven Indian sciaenids (Pisces: Perciformes, Sciaenidae) based on 16S rRNA and cytochrome c oxidase subunit I mitochondrial genes. *Molecular Biology Reports* 36(5): 831–839. <https://doi.org/10.1007/s11033-008-9252-1>

Lee WJ, Conroy J, Howell WH, Kocher TD (1995) Structure and evolution of teleost mitochondrial control regions. *Journal of Molecular Evolution* 41(1): 54–66. <https://doi.org/10.1007/BF00174041>

Liu L, Wang Z, Yanagimoto T, Gao T, Li P (2021) The taxonomic validity and phylogenetic relationships of genus *Platycephalus* Bloch, 1785 (Teleostei: Platycephalidae) in the Northwest Pacific inferred from the mitochondrial genome. *Thalassas: An International Journal of Marine Sciences* 37: 705–715. <https://doi.org/10.1007/s41208-021-00345-w>

Lowe TM, Eddy SR (1997) tRNAscan-SE: A program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Research* 25(5): 955–964. <https://doi.org/10.1093/nar/25.5.955>

Lunt DH, Whipple LE, Hyman BC (1998) Mitochondrial DNA variable number tandem repeats (VNTRs): utility and problems in molecular ecology. *Molecular Ecology* 7(11): 1441–1455. <https://doi.org/10.1046/j.1365-294x.1998.00495.x>

Mayfield JE, McKenna JF (1978) A-T rich sequences in vertebrate DNA. A possible explanation of q-banding in metaphase chromosomes. *Chromosoma* 67(2): 157–163. <https://doi.org/10.1007/BF00293173>

Meyer A (1993) Evolution of mitochondrial DNA in fishes. In: Hochachka PW, Mommsen TP (ed), *Biochemistry and molecular biology of fishes*. Elsevier Science Publishers, Amsterdam, 38 pp.

Mignotte F, Gueride M, Champagne AM, Mounolou JC (1990) Direct repeats in the non-coding region of rabbit mitochondrial DNA. Involvement in the generation of intra- and inter-individual heterogeneity. *European Journal of Biochemistry* 194(2): 561–571. <https://doi.org/10.1111/j.1432-1033.1990.tb15653.x>

Minhas BF, Beck EA, Cheng CC, Catchen J (2023) Novel mitochondrial genome rearrangements including duplications and extensive heteroplasmy could underlie temperature adaptations in Antarctic notothenioid fishes. *Scientific Reports* 13(1): 6939. <https://doi.org/10.1038/s41598-023-34237-1>

Miya M, Sato Y, Fukunaga T, Sado T, Poulsen JY, Sato K, Minamoto T, Yamamoto S, Yamakawa H, Araki H, Kondoh M, Iwasaki W (2015) MiFish, a set of universal PCR primers for metabarcoding environmental DNA from fishes: detection of more than 230 subtropical marine species. Royal Society Open Science 2(7): 150088 <https://doi.org/10.1098/rsos.150088>

Moeckel C, Zaravinos A, Georgakopoulos-Soares I (2023) Strand asymmetries across genomic processes. Computational and Structural Biotechnology Journal 21: 2036–2047. <https://doi.org/10.1016/j.csbj.2023.03.007>

Nalbant TT (1993) Some problems in the systematics of the genus *Cobitis* and its relatives (Pisces, Ostariophysi, Cobitidae). Revue Roumaine de Biologie: Serie Biologie Animale 38(2): 101–110.

Nicholls TJ, Minczuk M (2014) In D-loop: 40 years of mitochondrial 7S DNA. Experimental Gerontology 56: 175–181. <https://doi.org/10.1016/j.exger.2014.03.027>

Norman JA, Moritz C, Limpus CJ (1994) Mitochondrial DNA control region polymorphisms: genetic markers for ecological studies of marine turtles. Molecular Ecology 3(4): 363–373. <https://doi.org/10.1111/j.1365-294X.1994.tb00076.x>

Pereira SL (2000) Mitochondrial genome organization and vertebrate phylogenetics. Genetics and Molecular Biology 23(4): 745–752. <https://doi.org/10.1590/S1415-47572000000400008>

Pereira F, Soares P, Carneiro J, Pereira L, Richards MB, Samuels DC, Amorim A (2008) Evidence for variable selective pressures at a large secondary structure of the human mitochondrial DNA control region. Molecular Biology and Evolution 25(12): 2759–2770. <https://doi.org/10.1093/molbev/msn225>

Perna NT, Kocher TD (1995) Patterns of nucleotide composition at fourfold degenerate sites of animal mitochondrial genomes. Journal of Molecular Evolution 41(3): 353–358. <https://doi.org/10.1007/BF01215182>

Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. Systematic Biology 61(3): 539–542. <https://doi.org/10.1093/sysbio/sys029>

Saccone C, Attimonelli M, Sbisà E (1987) Structural elements highly preserved during the evolution of the D-loop-containing region in vertebrate mitochondrial DNA. Journal of Molecular Evolution 26(3): 205–211. <https://doi.org/10.1007/BF02099853>

Saitoh K, Sado T, Mayden RL, Hanzawa N, Nakamura K, Nishida M, Miya M (2006) Mitogenomic evolution and interrelationships of the Cypriniformes (Actinopterygii: Ostariophysi): the first evidence toward resolution of higherlevel relationships of the world's largest freshwater fish clade based on 59 whole mitogenome sequences. Journal of Molecular Evolution 63(6): 826–841. <https://doi.org/10.1007/s00239-005-0293-y>

Saitoh K, Chen WJ, Mayden RL (2010) Extensive hybridization and tetraployploidy in spined loach fish. Molecular Phylogenetics and Evolution 56(3): 1001–1010. <https://doi.org/10.1016/j.ympev.2010.04.021>

Sambrook J, Russell DW (2001) Molecular cloning: a laboratory manual. Cold Spring Harbor Press, New York, 58 pp.

Sarri C, Stamatidis C, Sarafidou T, Galara I, Godosopoulos V, Kolovos M, Liakou C, Tatsoglou S, Mamuris Z (2014) A new set of 16S rRNA universal primers for identification of animal species. Food Control 43: 35–41. <https://doi.org/10.1016/j.foodcont.2014.02.036>

Satoh TP, Miya M, Mabuchi K, Nishida M (2016) Structure and variation of the mitochondrial genome of fishes. BMC Genomics 17(1): 719. <https://doi.org/10.1186/s12864-016-3054-y>

Sawada Y (1982) Phylogeny and zoogeography of the superfamily Cobitoidea (Cyprinoidae, Cypriniformes). Memoirs of the Faculty of Fisheries, Hokkaido University 28: 65–223. <http://eprints.lib.hokudai.ac.jp/dspace/bitstream/2115/21871/1/2>

Shadel GS, Clayton DA (1997) Mitochondrial DNA maintenance in vertebrates. Annual Review of Biochemistry 66: 409–435. <https://doi.org/10.1146/annurev.biochem.66.1.409>

Sharp PM, Li WH (1986) An evolutionary perspective on synonymous codon usage in unicellular organisms. Journal of Molecular Evolution 24(1–2): 28–38. <https://doi.org/10.1007/BF02099948>

Shen Y, Wang J, Zhang F (2020) Complete mitochondrial genome of *Parabotia bimaculata* (Cypriniformes: Cobitidae: Botiinae), an endemic riverine loach in China and phylogenetic analysis for Botiinae. Thalassas: An International Journal of Marine Sciences 36: 387–393. <https://doi.org/10.1007/s41208-020-00200-4>

Singh TR, Shneor O, Huchon D (2008) Bird mitochondrial gene order: insight from 3 warbler mitochondrial genomes. Molecular Biology and Evolution 25(3): 475–477. <https://doi.org/10.1093/molbev/msn003>

Slechtová V, Bohlen J, Perdices A (2008) Molecular phylogeny of the freshwater fish family Cobitidae (Cypriniformes: Teleostei): delimitation of genera, mitochondrial introgression and evolution of sexual dimorphism. Molecular Phylogenetics and Evolution 47(2): 812–831. <https://doi.org/10.1016/j.ympev.2007.12.018>

Smith HM (1931) Descriptions of new genera and species of Siamese fishes. Proceedings of the United States National Museum: 79(2873): 1–48. <https://doi.org/10.5479/si.00963801.79-2873.1>

Stamatakis A (2006) RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. Bioinformatics (Oxford, England) 22(21): 2688–2690. <https://doi.org/10.1093/bioinformatics/btl446>

Sudasinghe H, Dahanukar N, Raghavan R, Ranasinghe T, Wijesooriya K, Pethiyagoda R, Rüber L, Meegaskumbura M (2024) The loach genus *Lepidocephalichthys* (Teleostei: Cobitidae) in Sri Lanka and peninsular India: multiple colonizations and unexpected species diversity. Hydrobiologia 851: 1113–1133. <https://doi.org/10.1007/s10750-023-05321-4>

Sureandiran B, Karuppasamy K, Sundaramanickam A, Badhul Haq MA, Arumugam U, Ajith Kumar TT (2023) First record and DNA barcode of bigeye numbfish, *Narcine Oculifera* Carvalho, Compagno & Mee, 2002 (Elasmobranchii: Torpediniformes: Narcinidae) from Indian coastal waters. Thalassas: An International Journal of Marine Sciences 39: 1215–1221. <https://doi.org/10.1007/s41208-023-00569-y>

Taanman JW (1999) The mitochondrial genome: structure, transcription, translation and replication. Biochimica et Biophysica Acta 1410(2): 103–123. [https://doi.org/10.1016/S0005-2728\(98\)00161-3](https://doi.org/10.1016/S0005-2728(98)00161-3)

Tang Q, Liu H, Mayden R, Xiong B (2005) Comparison of evolutionary rates in the mitochondrial DNA cytochrome *b* gene and control region and their implications for phylogeny of the Cobitoidea (Teleostei: Cypriniformes). Molecular Phylogenetics and Evolution 39(2): 347–357. <https://doi.org/10.1016/j.ympev.2005.08.007>

Tomita K, Ueda T, Watanabe K (1996) RNA editing in the acceptor stem of squid mitochondrial tRNATyr. Nucleic Acids Research 24(24): 4987–4991. <https://doi.org/10.1093/nar/24.24.4987>

Trif N, Bordeianu M, Codrea VA (2022) A Maeotian (Late Miocene) freshwater fish-fauna from Romania. Palaeoworld 31(1): 140–152. <https://doi.org/10.1016/j.palwor.2021.01.004>

Valenciennes A, Cuvier G (1846) *Histoire naturelle des poissons*. Bertrand, Paris, 520–553 pp.

Wang Y, Guo R, Li H, Zhang X, Du J, Song Z (2011) The complete mitochondrial genome of the Sichuan taimen (*Hucho bleekeri*): Repetitive sequences in the control region and phylogenetic implications for Salmonidae. *Marine Genomics* 4(3): 221–228. <https://doi.org/10.1016/j.margen.2011.06.003>

Wang Y, Wen H, Yao J, Sun K, Wang W, Liu H, Yang D, Zhang F, Xiong F (2018) Complete mitochondrial genome of the Salangid icefish *Neosalanx taihuensis* (Actinopterygii: Osmeriformes: Salangidae). *Mitochondrial DNA. Part B, Resources* 3(2): 1040–1041. <https://doi.org/10.1080/23802359.2018.1511840>

Wang IC, Lin HD, Liang CM, Huang CC, Wang RD, Yang JQ, Wang WK (2020) Complete mitochondrial genome of the freshwater fish *Onychostoma lepturum* (Teleostei, Cyprinidae): genome characterization and phylogenetic analysis. *ZooKeys* 1005: 57–72. <https://doi.org/10.3897/zookeys.1005.57592>

Wang Y, Wen H, Zhai D, Liu H, Xiong F (2021) DNA barcoding for identification of fishes in Xiangjiaba reservoir area in the downstream section of the Jinsha River. *Conservation Genetics Resources* 13: 201–208. <https://doi.org/10.1007/s12686-021-01196-6>

Watanabe Y, Suematsu T, Ohtsuki T (2014) Losing the stem-loop structure from metazoan mitochondrial tRNAs and co-evolution of interacting factors. *Frontiers in Genetics* 5: 109. <https://doi.org/10.3389/fgene.2014.00109>

Wen ZY, Xie BW, Qin CJ, Wang J, Deng YY, Li R, Zou YC (2017) The complete mitochondrial genome of a threatened loach (*Beaufortia kweichowensis*) and its phylogeny. *Conservation Genetics Resources* 9: 565–568. <https://doi.org/10.1007/s12686-017-0723-3>

Wu HW, Lin RD, Chen QX, Chen XL, He MQ (1977) *Zhongguo like yulei zhi*. People's Press, Shanghai, 229–394 pp.

Wu J, He Y, Ren H, Zhang Y, Du Z, Xie M, Zhu G, Wang Q, Jiang Y, He T, Wen A (2016) The complete mitochondrial genome sequence of *Beaufortia szechuanensis* (Cypriniformes, Balitoridae). *Mitochondrial DNA. Part A, DNA Mapping, Sequencing, and Analysis* 27(4): 2535–2536. <https://doi.org/10.3109/19401736.2015.1038792>

Xing ZP, Ling X, Wang X, Hu HY, Huang YX (2022) Novel gene rearrangement pattern in mitochondrial genome of *Ooencyrtus plautus* Huang & Noyes, 1994: new gene order in Encyrtidae (Hymenoptera, Chalcidoidea). *ZooKeys* 1124: 1–21. <https://doi.org/10.3897/zookeys.1124.83811>

Xu DM, Yang Z, Xu N, Xiong MH, Lei P, Que YF (2016) The complete mitochondrial genome of *Lepturichthys fimbriata* (Teleostei, Balitoridae, Balitorinae). *Mitochondrial DNA. Part A, DNA Mapping, Sequencing, and Analysis* 27: 1064–1065. <https://doi.org/10.3109/19401736.2014.928869>

Xu ZG, Wu L, Hua S, Hua C, Huang T, Zhao Y (2021) Structural variation and phylogenetic relationship of *Geospiza magnirostris* based on mitochondrial control region. *Biologia* 76: 1367–1373. <https://doi.org/10.2478/s11756-020-00669-7>

Yu P, Ding S, Yang Q, Li X, Wan Q (2016) The complete mitochondrial genome of *Sinibotia robusta* (Cypriniformes: Cobitidae). *Mitochondrial DNA. Part A, DNA Mapping, Sequencing, and Analysis* 27(5): 3471–3472. <https://doi.org/10.3109/19401736.2015.1066353>

Yu P, Zhou L, Yang WT, Miao LJ, Li Z, Zhang XJ, Wang Y, Gui JF (2021) Comparative mitogenome analyses uncover mitogenome features and phylogenetic implications of the subfamily Cobitinae. *BMC Genomics* 22(1): 50. <https://doi.org/10.1186/s12864-020-07360-w>

Zhang F, Shen Y (2019) Characterization of the complete mitochondrial genome of *Rhinogobius leavelli* (Perciformes: Gobiidae: Gobionellinae) and its phylogenetic analysis for Gobionellinae. *Biologia* 74: 493–499. <https://doi.org/10.2478/s11756-018-00189-5>

Zhang C, Ding C, Ding L, Chen L, Hu J, Tao J, Jiang X (2019) Large-scale cascaded dam constructions drive taxonomic and phylogenetic differentiation of fish fauna in the Lancang River, China. *Reviews in Fish Biology and Fisheries* 29: 895–916. <https://doi.org/10.1007/s11160-019-09580-0>

Zhang D, Gao F, Jakovlić I, Zou H, Zhang J, Li WX, Wang GT (2020) PhyloSuite: An integrated and scalable desktop platform for streamlined molecular sequence data management and evolutionary phylogenetics studies. *Molecular Ecology Resources* 20(1): 348–355. <https://doi.org/10.1111/1755-0998.13096>

Zhang M, Zhou Q, Xiang H, Wang J, Lan X, Luo Q, Jiang W (2023) Complete mitochondrial genome of *Rectoris luxiensis* (Teleostei, Cyprinidae): characterisation and phylogenetic implications. *Biodiversity Data Journal* 11: e96066. <https://doi.org/10.3897/BDJ.11.e96066>

Zhong D, Zhao GJ, Zhang ZS, Xun AL (2002) Advance in the entire balance and local unbalance of base distribution in genome. *Hereditas (Beijing)* 24(3): 351–355. [in Chinese]

Zhou X, Yu Y, Li Y, Wu J, Zhang X, Guo X, Wang W (2014) Comparative analysis of mitochondrial genomes in distinct nuclear ploidy loach *Misgurnus anguillicaudatus* and its implications for polyploidy evolution. *PLoS ONE* 9(3): e92033. <https://doi.org/10.1371/journal.pone.0092033>

Zou YC, Xie BW, Qin CJ, Wang YM, Yuan DY, Li R, Wen ZY (2017) The complete mitochondrial genome of a threatened loach (*Sinibotia reevesae*) and its phylogeny. *Genes & Genomics* 39: 767–778. <https://doi.org/10.1007/s13258-017-0541-8>

Zou YC, Chen M, Qin CJ, Wang YM, Li R, Qi ZM, Wen ZY (2018) Identification of the complete mitochondrial genome of *Garra pingi pingi* (Cypriniformes, Cyprinidae). *Conservation Genetics Resources* 10: 693–696. <https://doi.org/10.1007/s12686-017-0903-1>

Zou Y, Liu T, Li Q, Wen Z, Qin C, Li R, Wang D (2019) Complete mitochondrial genome of *Hemiculter tchangi* (Cypriniformes, Cyprinidae). *Conservation Genetics Resources* 11: 1–4. <https://doi.org/10.1007/s12686-017-0949-0>

## Supplementary material 1

### Supplementary files

Authors: Min Zhou, Cheng Wang, Ziyue Xu, Zhicun Peng, Yang He, Ying Wang

Data type: docx

Explanation note: **figure S1.** Images of biological sample for this study. **table S1.** Primers used for amplification of the mitochondrial genome of *Lepidocephalichthys berdmorei*. **table S2.** Species and GenBank accession numbers of mitogenomes used in this study. **table S3.** Nucleotide contents of genes and the mitochondrial genome skew of *Lepidocephalichthys berdmorei*.

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: <https://doi.org/10.3897/zookeys.1221.129136.suppl1>